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Detection of copy number aberrations in clinical diagnostics has been limited to few genes in MLPA and low resolution in microarrays. We have developed and validated a low-coverage whole genome sequencing assay for genome-wide and high-resolution detection of copy number aberrations (CNAs) from inherited disorders.

**a**

raw read counts

598k x 5 kbp

$E = 0.165, \delta_E = 0.178$

log read count

chromosome

**b**

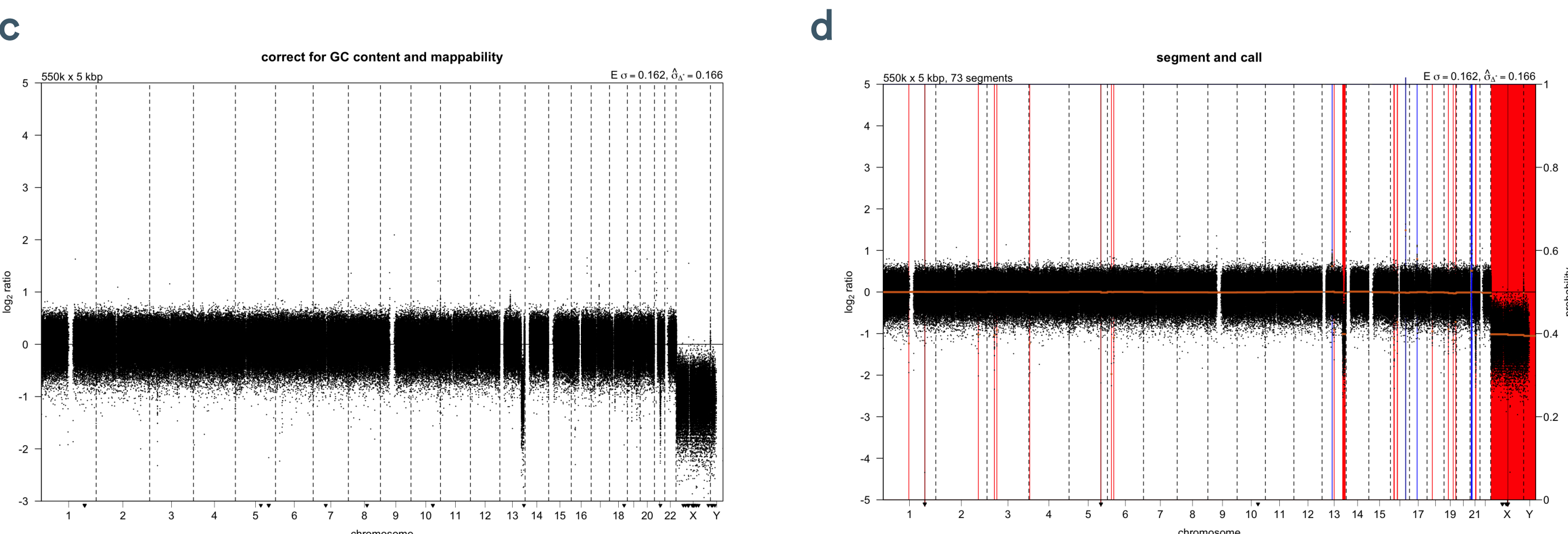
filter problematic areas

598k x 5 kbp

$E = 0.165, \delta_E = 0.178$

log read count

chromosome



**Figure 1** Methods for detecting copy number aberrations using low-coverage whole genome sequencing. a Sequencing and binning, b filtering, c GC content and mappability and d segmentation and calling.

Quantitative PCR is applied to confirm pathogenic and likely pathogenic Del/Dup variants from probands as well as for screening of disease-causing mutations in the family members. Validation of Del/Dup confirmation assay was performed using 15 well-characterized reference samples and 32 Del/Dup assays (**Table 3**).

**Table 3** Analytic validation of quantitative PCR assay for CNA confirmation.

Performance metric	Value	Measurements
Sensitivity	0.954	TP: 21
Specificity	0.920	TN: 23
Positive predictive value	0.913	FP: 2
Accuracy	0.936	FN:1
Repeatability	1.000	
Reproducibility	1.000	

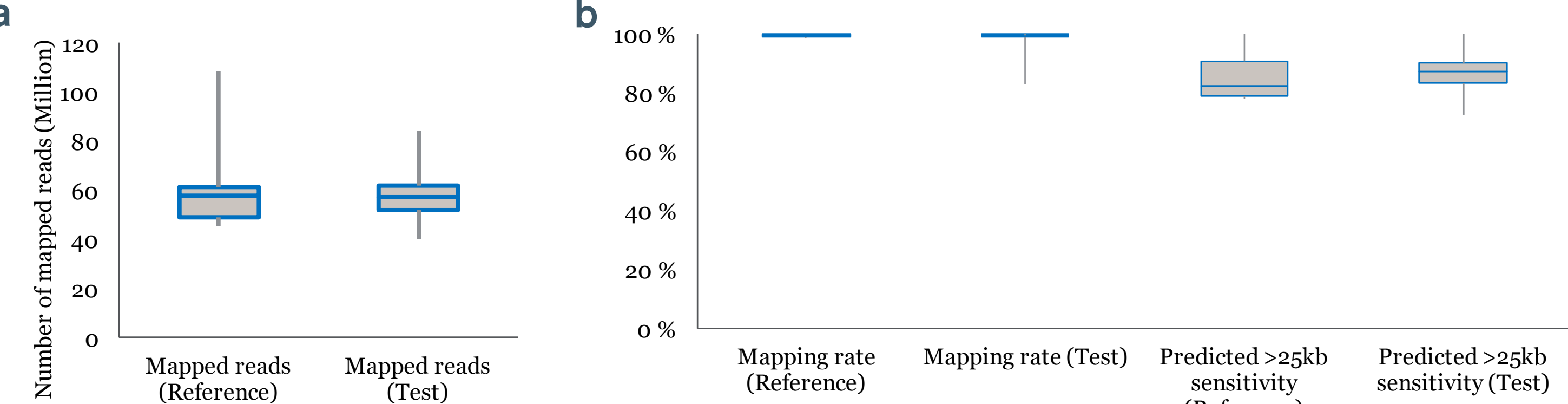
A well-characterized golden-standard reference sample (NA12878) is added into each sample processing, sequencing and data analysis batch to control for quality of the testing and to assess the sensitivity of the performed assays (**Table 4**). For each clinical sample and in-process quality control sample, number of mapped reads, mapping rate and predicted >25kb are measured (**Figure 4**) to ensure that each analysis fulfills the criteria derived from the analytic validation studies.

#### Table 4 Quality metrics and acceptance criteria for clinical testing

Quality metrics	Acceptance criteria	Actual*
n-process reference sample sensitivity (>50 kb)	>0.8	1.0
Number of mapped reads	>39M	58.2M

Number of mapped reads (Million)

Dataset	Min	Q1	Median	Q3	Max
Mapped reads (Reference)	~4.5	~5.0	~5.5	~6.0	~11.0
Mapped reads (Test)	~4.0	~5.5	~6.0	~6.5	~8.5



**Figure 4** In-process quality control of low-coverage CNA detection during a 3-month period. **a** Number of mapped reads, **b** mapping rate and predicted >25kb sensitivity are measured for reference samples and test samples.

Blueprint Genetics performs analytic validation of all laboratory and data analysis assays per ACMG guidelines (1). Metrics included in the analytic validation of Blueprint Genetics diagnostics assays are described in **Table 1**.

**Table 1** Metrics applied in the analytic validation of CNAs in inherited disorders.

**PRECISION** is a statistical measure of the performance of the assay to generate correct test result. For estimation of the precision, true positives (TP), true negatives (TN), false positives (FP) and false negatives (FN) are measured and sensitivity, specificity, positive predictive value and accuracy are calculated.

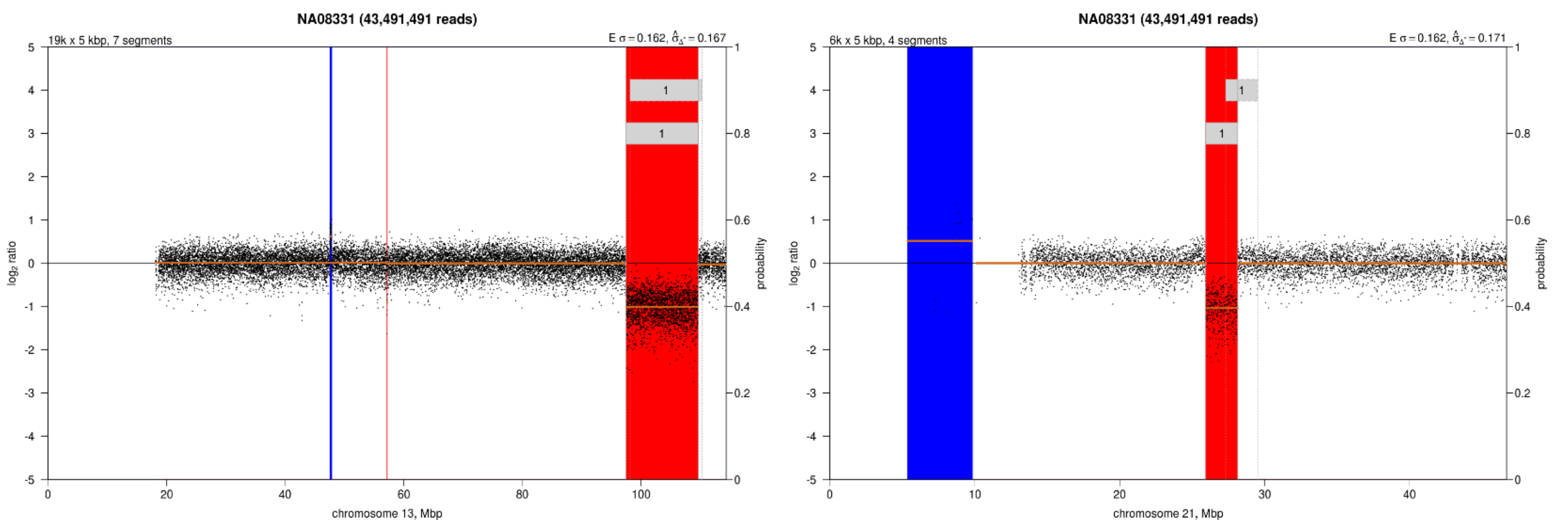
**CLINICAL SENSITIVITY** provides a statistical measure of the assays clinical performance. It reflects the assay's ability to provide a diagnosis in specific clinical use cases. Clinical sensitivity is measured by calculating the extent in which all clinical diagnosis scenarios are met.

**REPORTABLE RANGE** is the functional range of an assay over which the analyte can be analyzed.

**REPEATABILITY** is the technical variation in measurements taken by a single person on the same instrument, on the same experiment, under the same conditions, and in a short time.

**REPRODUCIBILITY** is the ability of a test result to be duplicated under all variable conditions (different users, between reagent lots, using different instruments and different testing times).

Blueprint Genetics applies independent, publicly available sample materials and data sets as the reference materials in all validation studies to ensure full traceability of the validation results. The CNA calls from the reference samples were compared to Affymetrix Genome-Wide Human SNP Array 6.0, G-banded karyotyping analysis and fluorescence in situ hybridization (FISH) results (**Figure 2**). 28 reference samples with 34 confirmed chromosomal aberrations and the golden standard reference sample (NA12878) were applied in the validation (**Table 2**). Sensitivity to detect >25kb Del/Dups was estimated based on diversity of the sequencing library (**Figure 3**).



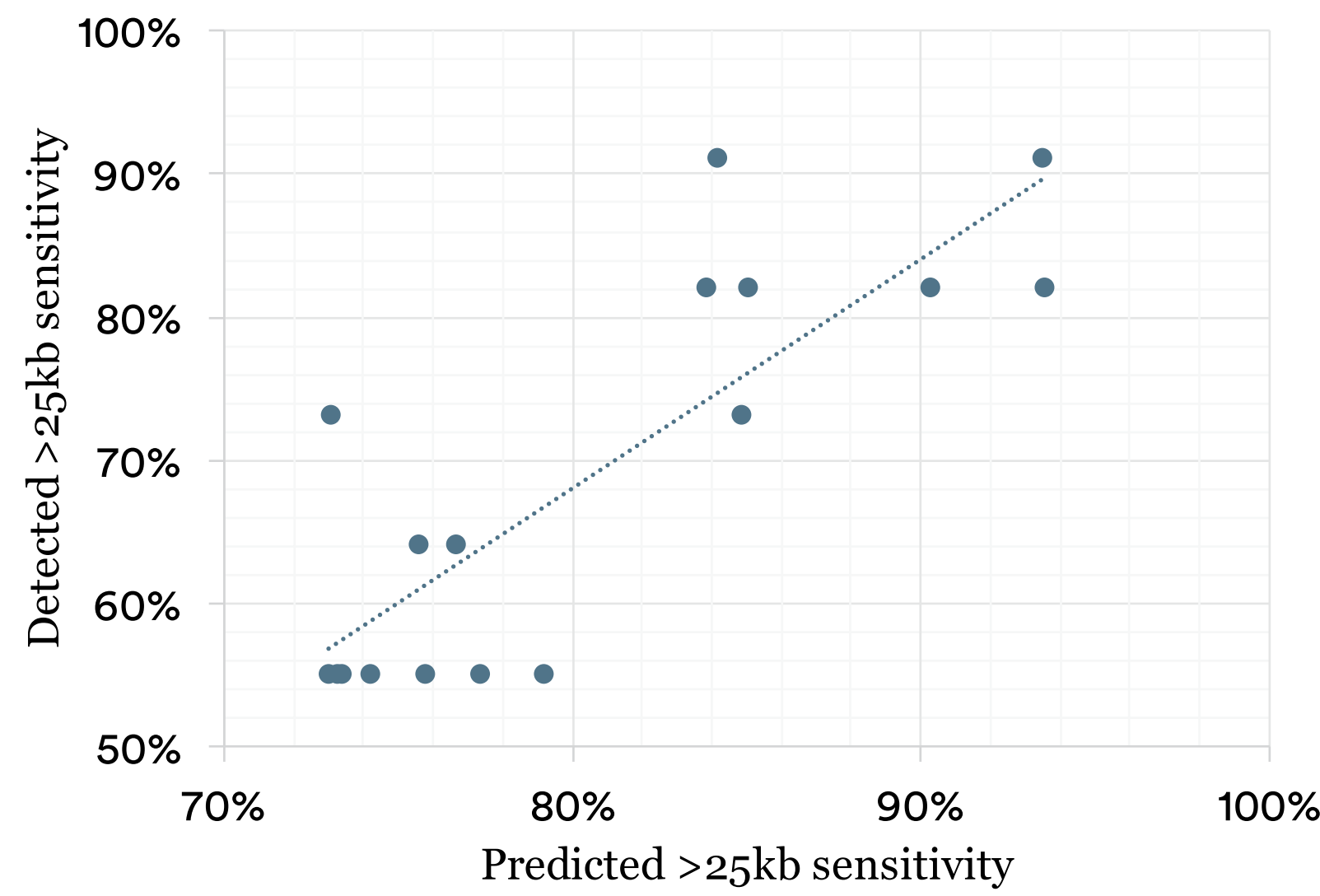
**Figure 2** Known deletions in chromosomes 13 and 21. CNA calls aligned with microarray data showing 1 copy in different genome coordinate systems (hg19/hg39). Red bars depict deletions and blue bars depict duplications.

### Table 2 Analytic validation of detection of CNAs.

Performance metric	Value	Measurements
Sensitivity (chromosomal aberrations, >100 kb)	0.97	TP/FN: 33 / 1
Sensitivity (Del/Dups)		
10-25 kb	0.06	TP/FN: 16 / 273
25-50 kb	0.76	TP/FN: 143 / 44
>50 kb	0.99	TP/FN: 101 / 1
Clinical sensitivity (% genes that are analyzed*)		130 284 out of 138 259 segments
Clinical sensitivity (% genome that is analyzed**)	94.2 %	used
	85.2 %	527 553 out of 619 150 segments
		used
Reportable range	10 kb and larger	
Repeatability	0.86	
Reproducibility	0.85	

\* Coding regions of the human genes contain regions that are affected by pseudogenes, repeats and extreme GC-content that are not reproducibly analyzed using short read sequencing.

\*\* Human genome contains difficult to sequence regions such as centromeres and telomeres.



**Figure 3** Correlation ( $R^2=0.68$ ) between predicted and detected >25kb Del/Dup sensitivity.

(1) Rehm HL, Bale SJ, Bayrak-Toydemir P, Berg JS, Brown KK, Deignan JL, Friez MJ, Funke BH, Hegde MR, Lyon E; Working Group of the American College of Medical Genetics and Genomics Laboratory Quality Assurance Committee. ACMG clinical laboratory standards for next-generation sequencing. *Genet Med*. 2013 Sep;15(9):733-47