

Mitochondrial DNA Depletion Syndrome Panel

Test code: ME0201

In addition, it also includes the maternally inherited mitochondrial genome. Is ideal for patients with a clinical suspicion of mitochondrial DNA depletion syndrome. The genes on this panel are included in the Comprehensive Metabolism Panel.

Mitochondrial DNA depletion syndrome (MDS) is a group of autosomal recessive syndromes that are clinically and genetically heterogeneous. This panel can be used in differential diagnosis and it is also part of the Comprehensive Metabolism Panel.

About Mitochondrial DNA Depletion Syndrome

Mitochondrial DNA depletion syndrome (MDS) refers to a group of disorders which cause the affected tissues to suffer from a significant drop in mitochondrial DNA. Symptoms may manifest as myopathic (mutations in *TK2*, *RRM2B* and *AG*), encephalomyopathic (*SUCLA2*, *SUCLG1*, and *RRM2B*) hepatocerebral (*DGUOK*, *MPV17*, *POLG* and *C10ORF2*), and/or neurogastrointestinal (*TYMP*). These syndromes affect tissue found in the muscle, liver, or both the muscle and brain, respectively. Typically, the condition is fatal in infancy or early childhood. However some patients have survived into puberty with a myopathic variant and some into adulthood with a *SUCLA2* encephalomyopathic variant. There are some preliminary treatments that have shown to reduce symptoms but currently no curative treatment for any form of MDSs is available. Nuclear encoded genes associated with multiple mtDNA deletions or mtDNA depletion include *POLG*, *POLG2*, *C10orf2*, *SLC25A4* (autosomal progressive external ophthalmoplegia), *TYMP* (mitochondrial neurogastrointestinal encephalomyopathy), *POLG* (Alpers-Huttenlocher syndrome), *POLG*, *C10orf2*, *OPA1* (ataxia neuropathy syndromes 2), *TK2* (infantile myopathy / spinal muscular atrophy), *DGUOK* (encephalomyopathy and liver failure), *SUCLA2* (hypotonia, movement disorder, and/or Leigh syndrome with methylmalonic aciduria) and *RRM2B* (hypotonia, encephalopathy, renal tubulopathy, lactic acidosis).

Availability

Results in 3-4 weeks

Gene set description

Genes in the Mitochondrial DNA Depletion Syndrome Panel and their clinical significance

| Gene | Associated phenotypes | Inheritance | ClinVar | HGMD |
|----------------------|---|-------------|---------|------|
| AGK* | Sengers syndrome, Cataract 38 | AR | 18 | 27 |
| APTX | Ataxia, early-onset, with oculomotor apraxia and hypoalbuminemia | AR | 14 | 46 |
| AUH | 3-methylglutaconic aciduria | AR | 12 | 11 |
| C10ORF2 | Perrault syndrome, Mitochondrial DNA depletion syndrome, Progressive external ophthalmoplegia with mitochondrial DNA deletions, autosomal dominant, 3 | AR | 37 | 80 |
| C12ORF65 | Spastic paraplegia, Combined oxidative phosphorylation deficiency | AR | 10 | 11 |
| DGUOK | Mitochondrial DNA depletion syndrome, Portal hypertension, noncirrhotic, Progressive external ophthalmoplegia with mitochondrial DNA deletions, autosomal recessive 4 | AR | 23 | 62 |
| FBXL4 | Mitochondrial DNA depletion syndrome | AR | 55 | 47 |
| MFN2 | Hereditary motor and sensory neuropathy, Charcot-Marie-Tooth disease | AD/AR | 70 | 223 |

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|---------|--|---------------|----|----|
| MPV17 | Mitochondrial DNA depletion syndrome | AR | 35 | 50 |
| MT-ATP6 | Neuropathy, ataxia, and retinitis pigmentosa, Leber hereditary optic neuropathy, Ataxia and polyneuropathy, adult-onset, Cardiomyopathy, infantile hypertrophic, Leigh syndrome, Striatonigral degeneration, infantile, mitochondrial | Mitochondrial | 19 | |
| MT-ATP8 | Cardiomyopathy, apical hypertrophic, and neuropathy, Cardiomyopathy, infantile hypertrophic | Mitochondrial | 4 | |
| MT-CO1 | Myoglobinuria, recurrent, Leber hereditary optic neuropathy, Sideroblastic anemia, Cytochrome C oxidase deficiency | Mitochondrial | 17 | |
| MT-CO2 | Cytochrome c oxidase deficiency | Mitochondrial | 8 | |
| MT-CO3 | Cytochrome c oxidase deficiency, Leber hereditary optic neuropathy | Mitochondrial | 9 | |
| MT-CYB | Leber hereditary optic neuropathy | Mitochondrial | 69 | |
| MT-ND1 | Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes, Leber hereditary optic neuropathy, Leber optic atrophy and dystonia | Mitochondrial | 21 | |
| MT-ND2 | Leber hereditary optic neuropathy, Mitochondrial complex I deficiency | Mitochondrial | 6 | |
| MT-ND3 | Leber optic atrophy and dystonia, Mitochondrial complex I deficiency | Mitochondrial | 7 | |
| MT-ND4 | Leber hereditary optic neuropathy, Leber optic atrophy and dystonia, Mitochondrial complex I deficiency | Mitochondrial | 11 | |
| MT-ND4L | Leber hereditary optic neuropathy | Mitochondrial | 2 | |
| MT-ND5 | Myoclonic epilepsy with ragged red fibers, Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes, Leber hereditary optic neuropathy, Mitochondrial complex I deficiency | Mitochondrial | 19 | |
| MT-ND6 | Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes, Oncocytoma, Leber hereditary optic neuropathy, Leber optic atrophy and dystonia, Mitochondrial complex I deficiency | Mitochondrial | 16 | |
| MT-RNR1 | Deafness, mitochondrial | Mitochondrial | 3 | |
| MT-RNR2 | Chloramphenicol toxicity/resistance | Mitochondrial | 2 | |
| MT-TA | Leber hereditary optic neuropathy, Mitochondrial multisystemic disorder, Progressive external ophthalmoplegia, Dilated cardiomyopathy (DCM) | Mitochondrial | 4 | |
| MT-TC | Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes | Mitochondrial | 3 | |
| MT-TD | Mitochondrial multisystemic disorder | Mitochondrial | 1 | |
| MT-TE | Diabetes-deafness syndrome, Mitochondrial myopathy, infantile, transient, Mitochondrial myopathy with diabetes | Mitochondrial | 5 | |
| MT-TF | Myoclonic epilepsy with ragged red fibers, Nephropathy, tubulointerstitial, Encephalopathy, mitochondrial, Epilepsy, mitochondrial, Myopathy, mitochondrial, Mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes | Mitochondrial | 7 | |
| MT-TG | Hypertrophic cardiomyopathy, Encephalopathy, Myopathy | Mitochondrial | 3 | |

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|---------|--|---------------|----|-----|
| MT-TH | Mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes | Mitochondrial | 4 | |
| MT-TI | Progressive external ophthalmoplegia | Mitochondrial | 7 | |
| MT-TK | Myoclonic epilepsy with ragged red fibers | Mitochondrial | 5 | |
| MT-TL1 | Cytochrome c oxidase deficiency, Myoclonic epilepsy with ragged red fibers, Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes, Diabetes-deafness syndrome, Cyclic vomiting syndrome, SIDS, susceptibility to | Mitochondrial | 14 | |
| MT-TL2 | Progressive external ophthalmoplegia, Mitochondrial multisystemic disorder | Mitochondrial | 5 | |
| MT-TM | Mitochondrial Myopathy, Leigh syndrome, Mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes | Mitochondrial | 1 | |
| MT-TN | Progressive external ophthalmoplegia | Mitochondrial | 3 | |
| MT-TP | Mitochondrial multisystemic disorder | Mitochondrial | 2 | |
| MT-TQ | Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes, Encephalopathy | Mitochondrial | 2 | |
| MT-TR | Dilated cardiomyopathy (DCM) | Mitochondrial | 2 | |
| MT-TS1 | Myoclonic epilepsy with ragged red fibers, Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes | Mitochondrial | 10 | |
| MT-TS2 | Mitochondrial multisystemic disorder | Mitochondrial | 2 | |
| MT-TT | | Mitochondrial | 5 | |
| MT-TV | Mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes | Mitochondrial | 3 | |
| MT-TW | Leigh syndrome, Mitochondrial Myopathy | Mitochondrial | 8 | |
| MT-TY | | Mitochondrial | 4 | |
| NDUFS1 | Mitochondrial complex I deficiency | AR | 22 | 28 |
| OPA1 | Optic atrophy, Optic atrophy 1, Optic atrophy with or without deafness, Ophthalmoplegia, myopathy, ataxia, and neuropathy, Behr syndrome, Mitochondrial DNA depletion syndrome 14 | AD/AR | 96 | 390 |
| OPA3 | Optic atrophy, 3-methylglutaconic aciduria | AD/AR | 13 | 15 |
| POLG | POLG-related ataxia neuropathy spectrum disorders, Sensory ataxia, dysarthria, and ophthalmoparesis, Alpers syndrome, Progressive external ophthalmoplegia with mitochondrial DNA deletions, Mitochondrial DNA depletion syndrome | AD/AR | 89 | 290 |
| POLG2 | Progressive external ophthalmoplegia with mitochondrial DNA deletions | AD | 5 | 14 |
| RRM2B | Progressive external ophthalmoplegia with mitochondrial DNA deletions, Mitochondrial DNA depletion syndrome | AD/AR | 41 | 41 |
| SLC25A3 | Mitochondrial phosphate carrier deficiency | AR | 2 | 5 |

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|-------------------------|--|-------|----|-----|
| SLC25A4 | Progressive external ophthalmoplegia with mitochondrial DNA deletions, Mitochondrial DNA depletion syndrome | AD/AR | 12 | 14 |
| SPG7 | Spastic paraplegia | AR | 69 | 111 |
| SUCLA2 | Mitochondrial DNA depletion syndrome | AR | 9 | 29 |
| SUCLG1 | Mitochondrial DNA depletion syndrome | AR | 12 | 28 |
| TIMM8A* | Mohr-Tranebjaerg syndrome, Jensen syndrome, Opticoacoustic nerve atrophy with dementia | XL | 11 | 21 |
| TK2 | Mitochondrial DNA depletion syndrome | AR | 38 | 52 |
| TMEM126A | Optic atrophy | AR | 3 | 1 |
| TYMP | Mitochondrial DNA depletion syndrome | AR | 84 | 94 |
| WFS1 | Wolfram syndrome, Deafness, Wolfram-like syndrome, autosomal dominant, Deafness, autosomal dominant 6/14/38, Cataract 41 | AD/AR | 69 | 362 |

*Some regions of the gene are duplicated in the genome. [Read more.](#)

The gene has suboptimal coverage (means <90% of the gene's target nucleotides are covered at >20x with mapping quality score (MQ>20) reads), and/or the gene has exons listed under Test limitations section that are not included in the panel as they are not sufficiently covered with high quality sequence reads.

The sensitivity to detect variants may be limited in genes marked with an asterisk (*) or number sign (#)

Gene refers to the HGNC approved gene symbol; Inheritance refers to inheritance patterns such as autosomal dominant (AD), autosomal recessive (AR), mitochondrial (mi), X-linked (XL), X-linked dominant (XLD) and X-linked recessive (XLR); ClinVar refers to the number of variants in the gene classified as pathogenic or likely pathogenic in this database ([ClinVar](#)); HGMD refers to the number of variants with possible disease association in the gene listed in Human Gene Mutation Database ([HGMD](#)). The list of associated, gene specific phenotypes are generated from [CGD](#) or Mitomap databases.

Non-coding disease causing variants covered by the panel

| Gene | Genomic location HG19 | HGVS | RefSeq | RS-number |
|--------|-----------------------|--------------|-------------|-------------|
| DGUOK | Chr2:74177650 | c.444-62C>A | NM_080916.2 | |
| DGUOK | Chr2:74177701 | c.444-11C>G | NM_080916.2 | rs536746349 |
| OPA1 | Chr3:193334932 | c.449-34dupA | NM_130837.2 | |
| OPA1 | Chr3:193374829 | c.2179-40G>C | NM_130837.2 | |
| TIMM8A | ChrX:100601671 | c.133-23A>C | NM_004085.3 | rs869320666 |
| WFS1 | Chr4:6271704 | c.-43G>T | NM_006005.3 | |

Test Strengths

The strengths of this test include:

- CAP accredited laboratory
- CLIA-certified personnel performing clinical testing in a CLIA-certified laboratory

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- Powerful sequencing technologies, advanced target enrichment methods and precision bioinformatics pipelines ensure superior analytical performance
- Careful construction of clinically effective and scientifically justified gene panels
- Some of the panels include the whole mitochondrial genome (please see the Panel Content section)
- Our Nucleus online portal providing transparent and easy access to quality and performance data at the patient level
- Our publicly available analytic validation demonstrating complete details of test performance
- ~2,000 non-coding disease causing variants in our clinical grade NGS assay for panels (please see 'Non-coding disease causing variants covered by this panel' in the Panel Content section)
- Our rigorous variant classification scheme
- Our systematic clinical interpretation workflow using proprietary software enabling accurate and traceable processing of NGS data
- Our comprehensive clinical statements

Test Limitations

The following exons are not included in the panel as they are not sufficiently covered with high quality sequence reads: *TK2* (NM_001271934:3). Genes with suboptimal coverage in our assay are marked with number sign (#) and genes with partial, or whole gene, segmental duplications in the human genome are marked with an asterisk (*) if they overlap with the UCSC pseudogene regions. Gene is considered to have suboptimal coverage when >90% of the gene's target nucleotides are not covered at >20x with mapping quality score (MQ>20) reads. The technology may have limited sensitivity to detect variants in genes marked with these symbols (please see the Panel content table above).

This test does not detect the following:

- Complex inversions
- Gene conversions
- Balanced translocations
- Some of the panels include the whole mitochondrial genome but not all (please see the Panel Content section)
- Repeat expansion disorders unless specifically mentioned
- Non-coding variants deeper than ± 20 base pairs from exon-intron boundary unless otherwise indicated (please see above Panel Content / non-coding variants covered by the panel).

This test may not reliably detect the following:

- Low level mosaicism in nuclear genes (variant with a minor allele fraction of 14.6% is detected with 90% probability)
- Stretches of mononucleotide repeats
- Low level heteroplasmy in mtDNA (>90% are detected at 5% level)
- Indels larger than 50bp
- Single exon deletions or duplications
- Variants within pseudogene regions/duplicated segments
- Some disease causing variants present in mtDNA are not detectable from blood, thus post-mitotic tissue such as skeletal muscle may be required for establishing molecular diagnosis.

The sensitivity of this test may be reduced if DNA is extracted by a laboratory other than Blueprint Genetics.

For additional information, please refer to the Test performance section and see our Analytic Validation.

Test performance

The Blueprint Genetics mitochondrial DNA depletion syndrome panel covers classical genes associated with mitochondrial DNA depletion syndrome. The genes on the panel have been carefully selected based on scientific literature, mutation databases and our experience.

Our panels are sliced from our high-quality whole exome sequencing data. Please see our sequencing and detection performance table for different types of alterations at the whole exome level (Table).

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Assays have been validated for different starting materials including EDTA-blood, isolated DNA (no FFPE), saliva and dry blood spots (filter card) and all provide high-quality results. The diagnostic yield varies substantially depending on the assay used, referring healthcare professional, hospital and country. Blueprint Genetics' Plus Analysis (Seq+Del/Dup) maximizes the chance to find a molecular genetic diagnosis for your patient although Sequence Analysis or Del/Dup Analysis may be a cost-effective first line test if your patient's phenotype is suggestive of a specific mutation type.

The genes on the panel have been carefully selected based on scientific literature, mutation databases and our experience.

Our panels are sectioned from our high-quality, clinical grade NGS assay. Please see our sequencing and detection performance table for details regarding our ability to detect different types of alterations (Table).

| | Sensitivity % (TP/(TP+FN)) | Specificity % |
|---|----------------------------|---------------|
| Single nucleotide variants | 99.89% (99,153/99,266) | >99.9999% |
| Insertions, deletions and indels by sequence analysis | | |
| 1-10 bps | 96.9% (7,563/7,806) | >99.9999% |
| 11-50 bps | 99.13% (2,524/2,546) | >99.9999% |
| Copy number variants (exon level dels/dups) | | |
| 1 exon level deletion (heterozygous) | 100% (20/20) | NA |
| 1 exon level deletion (homozygous) | 100% (5/5) | NA |
| 1 exon level deletion (het or homo) | 100% (25/25) | NA |
| 2-7 exon level deletion (het or homo) | 100% (44/44) | NA |
| 1-9 exon level duplication (het or homo) | 75% (6/8) | NA |
| Simulated CNV detection | | |
| 5 exons level deletion/duplication | 98.7% | 100.00% |
| Microdeletion/-duplication sdrs (large CNVs, n=37) | | |
| Size range (0.1-47 Mb) | 100% (37/37) | |

The performance presented above reached by Blueprint Genetics high-quality, clinical grade NGS sequencing assay with the following coverage metrics

| | |
|---|--------|
| Mean sequencing depth | 143X |
| Nucleotides with >20x sequencing coverage (%) | 99.86% |

Performance of Blueprint Genetics Mitochondrial Sequencing Assay.

| | Sensitivity % (TP/(TP+FN)) | Specificity |
|---------------------------------------|----------------------------|-------------|
| ANALYTIC VALIDATION (NA samples; n=4) | | |
| Single nucleotide variants | | |

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|---|--------------------|---------|
| Heteroplasmic (45-100%) | 100.0% (50/50) | 100.0% |
| Heteroplasmic (35-45%) | 100.0% (87/87) | 100.0% |
| Heteroplasmic (25-35%) | 100.0% (73/73) | 100.0% |
| Heteroplasmic (15-25%) | 100.0% (77/77) | 100.0% |
| Heteroplasmic (10-15%) | 100.0% (74/74) | 100.0% |
| Heteroplasmic (5-10%) | 100.0% (3/3) | 100.0% |
| Heteroplasmic (<5%) | 50.0% (2/4) | 100.0% |
| CLINICAL VALIDATION (n=76 samples) | | |
| All types | | |
| Single nucleotide variants n=2084 SNVs | | |
| Heteroplasmic (45-100%) | 100.0% (1940/1940) | 100.0% |
| Heteroplasmic (35-45%) | 100.0% (4/4) | 100.0% |
| Heteroplasmic (25-35%) | 100.0% (3/3) | 100.0% |
| Heteroplasmic (15-25%) | 100.0% (3/3) | 100.0% |
| Heteroplasmic (10-15%) | 100.0% (9/9) | 100.0% |
| Heteroplasmic (5-10%) | 92.3%(12/13) | 99.98% |
| Heteroplasmic (<5%) | 88.7% (47/53) | 99.93% |
| Insertions and deletions by sequence analysis n=42 indels | | |
| Heteroplasmic (45-100%) 1-10bp | 100.0% (32/32) | 100.0% |
| Heteroplasmic (5-45%) 1-10bp | 100.0% (3/3) | 100.0% |
| Heteroplasmic (<5%) 1-10bp | 100.0% (5/5) | >0.9999 |
| SIMULATION DATA /(mitomap mutations) | | |
| Insertions, and deletions 1-24 bps by sequence analysis; n=17 | | |
| Homoplasmic (100%) 1-24bp | 100.0% (17/17) | 99.98% |
| Heteroplasmic (50%) | 100.0% (17/17) | 99.99% |
| Heteroplasmic (25%) | 100.0% (17/17) | 100.0% |
| Heteroplasmic (20%) | 100.0% (17/17) | 100.0% |
| Heteroplasmic (15%) | 100.0% (17/17) | 100.0% |
| Heteroplasmic (10%) | 94.1% (16/17) | 100.0% |
| Heteroplasmic (5%) | 94.1% (16/17) | 100.0% |
| Copy number variants (separate artificial mutations; n=1500) | | |



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|---|-----------------|-------------------|
| Homoplasmic (100%) 500 bp, 1kb, 5 kb | 100.0% | 100.0% |
| Heteroplasmic (50%) 500 bp, 1kb, 5 kb | 100.0% | 100.0% |
| Heteroplasmic (30%) 500 bp, 1kb, 5 kb | 100.0% | 100.0% |
| Heteroplasmic (20%) 500 bp, 1kb, 5 kb | 99.7% | 100.0% |
| Heteroplasmic (10%) 500 bp, 1kb, 5 kb | 99.0% | 100.0% |
| The performance presented above reached by following coverage metrics at assay level (n=66) | | |
| | Mean of medians | Median of medians |
| Mean sequencing depth MQ0 (clinical) | 18224X | 17366X |
| Nucleotides with >1000x MQ0 sequencing coverage (%) (clinical) | 100% | |
| rho zero cell line (=no mtDNA), mean sequencing depth | 12X | |

Bioinformatics

The target region for each gene includes coding exons and ± 20 base pairs from the exon-intron boundary. In addition, the panel includes non-coding variants if listed above (Non-coding variants covered by the panel). Some regions of the gene(s) may be removed from the panel if specifically mentioned in the "Test limitations" section above. The sequencing data generated in our laboratory is analyzed with our proprietary data analysis and annotation pipeline, integrating state-of-the-art algorithms and industry-standard software solutions. Incorporation of rigorous quality control steps throughout the workflow of the pipeline ensures the consistency, validity and accuracy of results. Our pipeline is streamlined to maximize sensitivity without sacrificing specificity. We have incorporated a number of reference population databases and mutation databases such as, but not limited to, [1000 Genomes Project](#), [gnomAD](#), [ClinVar](#) and [HGMD](#) into our clinical interpretation software to make the process effective and efficient. For missense variants, *in silico* variant prediction tools such as SIFT, PolyPhen, MutationTaster are used to assist with variant classification. Through our online ordering and statement reporting system, Nucleus, the customer has an access to details of the analysis, including patient specific sequencing metrics, a gene level coverage plot and a list of regions with inadequate coverage if present. This reflects our mission to build fully transparent diagnostics where customers have easy access to crucial details of the analysis process.

Clinical interpretation

We provide customers with the most comprehensive clinical report available on the market. Clinical interpretation requires a fundamental understanding of clinical genetics and genetic principles. At Blueprint Genetics, our PhD molecular geneticists, medical geneticists and clinical consultants prepare the clinical statement together by evaluating the identified variants in the context of the phenotypic information provided in the requisition form. Our goal is to provide clinically meaningful statements that are understandable for all medical professionals regardless of whether they have formal training in genetics.

Variant classification is the corner stone of clinical interpretation and resulting patient management decisions. Our classifications follow the [Blueprint Genetics Variant Classification Schemes](#) based on the [ACMG guideline 2015](#). Minor modifications were made to increase reproducibility of the variant classification and improve the clinical validity of the report. Our experience with tens of thousands of clinical cases analyzed at our laboratory allowed us to further develop the industry standard.

The final step in the analysis of sequence variants is confirmation of variants classified as pathogenic or likely pathogenic using bi-directional Sanger sequencing. Variant(s) fulfilling the following criteria are not Sanger confirmed: the variant quality score is above the internal threshold for a true positive call, and visual check-up of the variant at IGV is in-line with the variant call. Reported variants of uncertain significance are confirmed with bi-directional Sanger sequencing only if the quality score is below our internally defined quality score for true positive call. Reported copy number variations with a size <10 exons are confirmed by orthogonal methods such as qPCR if the specific CNV has been seen less than three times at Blueprint Genetics.



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Our clinical statement includes tables for sequencing and copy number variants that include basic variant information (genomic coordinates, HGVS nomenclature, zygosity, allele frequencies, in silico predictions, OMIM phenotypes and classification of the variant). In addition, the statement includes detailed descriptions of the variant, gene and phenotype(s) including the role of the specific gene in human disease, the mutation profile, information about the gene's variation in population cohorts and detailed information about related phenotypes. We also provide links to the references used, congress abstracts and mutation variant databases used to help our customers further evaluate the reported findings if desired. The conclusion summarizes all of the existing information and provides our rationale for the classification of the variant.

Identification of pathogenic or likely pathogenic variants in dominant disorders or their combinations in different alleles in recessive disorders are considered molecular confirmation of the clinical diagnosis. In these cases, family member testing can be used for risk stratification within the family. In the case of variants of uncertain significance (VUS), we do not recommend family member risk stratification based on the VUS result. Furthermore, in the case of VUS, we do not recommend the use of genetic information in patient management or genetic counseling.

Our interpretation team analyzes millions of variants from thousands of individuals with rare diseases. Thus, our database, and our understanding of variants and related phenotypes, is growing by leaps and bounds. Our laboratory is therefore well positioned to re-classify previously reported variants as new information becomes available. If a variant previously reported by Blueprint Genetics is re-classified, our laboratory will issue a follow-up statement to the original ordering health care provider at no additional cost.

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ICD codes

Commonly used ICD-10 codes when ordering the Mitochondrial DNA Depletion Syndrome Panel

| ICD-10 | Disease |
|---------|--|
| F84.2 | Rett syndrome |
| G71.3 | Mitochondrial DNA depletion syndrome |
| H49.40 | Progressive external ophthalmoplegia |
| G11.9 | Hereditary ataxia |
| C94.2 | Acute Megakaryoblastic Leukemia |
| K59.8 | Chronic Intestinal Pseudoobstruction |
| T36.5 | Adverse effect of aminoglycosides |
| G93.41 | Metabolic Encephalopathy |
| H49.81 | Kearns Sayre Syndrome |
| E88.42 | MERFF Syndrome |
| H47.013 | Nonarteritic Anterior Ischemic Optic Neuropathy |
| G60.2 | Neuropathy in association with hereditary ataxia |
| G30 | Alzheimer's Disease |
| G25.5 | Chorea |
| G40 | Epilepsy and recurrent seizures |

| | |
|--------|---|
| I42 | Cardiomyopathy |
| N26.9 | Focal Segmental Glomerulosclerosis |
| G31.82 | Leigh's Disease |
| H47.2 | Leber's hereditary optic neuropathy |
| G71.3 | Mitochondrial Myopathy |
| I42.1 | Hypertrophic Cardiomyopathy |
| E11.9 | Non-Insulin Dependent Diabetes Mellitus |
| Z86.74 | Personal history of sudden cardiac arrest |
| H90.3 | Sensorineural Hearing Loss |

Accepted sample types

- EDTA blood, min. 1 ml
- Purified DNA, min. 3µg*
- Saliva (Oragene DNA OG-500 kit)

Label the sample tube with your patient's name, date of birth and the date of sample collection.

Note that we do not accept DNA samples isolated from formalin-fixed paraffin-embedded (FFPE) tissue.

Resources

- [Alpers Awareness](#)
- [Alpers Awareness](#)
- [GeneReviews - Deoxyguanosine Kinase Deficiency](#)
- [GeneReviews - Mitochondrial Neurogastrointestinal Encephalomyopathy](#)
- [GeneReviews - POLG-Related Disorders](#)
- [GeneReviews - RRM2B-Related Mitochondrial Disease](#)
- [GeneReviews - SUCLA2 Deficiency](#)
- [GeneReviews - TK2-Related Mitochondrial DNA Depletion Myopathy](#)
- [GeneReviews- *RRM2B*-Related Mitochondrial Disease](#)
- [GeneReviews- *SUCLA2*-Related Mitochondrial DNA Depletion Syndrome, Encephalomyopathic Form with Methylmalonic Aciduria](#)
- [GeneReviews- Deoxyguanosine Kinase Deficiency](#)
- [GeneReviews- K2-Related Mitochondrial DNA Depletion Syndrome, Myopathic Form](#)
- [GeneReviews- Mitochondrial Neurogastrointestinal Encephalopathy Disease](#)
- [GeneReviews- POLG-Related Disorders.](#)
- [NORD - Alpers-Huttenlocher syndrome](#)
- [NORD - Mitochondrial Neurogastrointestinal Encephalomyopathy](#)
- [United Mitochondrial Disease Foundation](#)